



SERINE PROTEASE INHIBITOR

Field of the Invention

This invention relates to a serine protease inhibitor. The inhibitor contains a substance which binds to the substrate of a serine protease in competition therewith.

Background

Enzymes are proteinaceous, macromolecular organic catalysis produced by biogenic cells. Serine proteases are enzymes which in which the hydroxyl group of a serine residue is the active center. Examples of such serine proteases include: pepsin, trypsin, and chymotrypsin, which serve as digestive enzymes for animals; acrosin, which is capable of inducing lytic dissolution of the hyaline layer of an egg during the course of fecundation; thrombin, plasmin, activated blood coagulation factor II, activated blood coagulation factor VII, activated blood coagulation factor IX, activated blood coagulation factor X, activated blood coagulation factor XI, and activated blood coagulation factor XII, which participate in the coagulation and adenolysis of blood; and elastases.

The activated blood coagulating factors, including thrombin, are so deeply related to blood coagulation that the anomalous sthenia of their activities leads directly to disseminates intravascular coagulation (DIC) and other similar abnormal phenomena of blood coagulation. Thrombin acts on blood platelets and promotes the coagulation thereof. Though plasmin, uPA (urkinase), and tPA (tissue plasminogen activating factor) fulfill the function of decomposing fibrin in relation to adenolysis, their states of active sthenia form a cause

The serine proteases are deeply related to such various diseases as are implied above. In the therapy of these diseases, the inhibition of serine protease activity is very important.

The center of the catalyst for a serine protease is invariably composed of an active serine residue, the imidazole ring of a histidine residue existing in the proximity thereof, and the β -carboxyl group electric charge relay system of an aspartic acid residue. Though the substrate varies with the species of protease, the hydrolysis reaction with a protease is made to proceed by the reaction of the acyl moiety (-CO) of the substrate molecule (CO-NH) with the hydroxyl group of an active serine. In nature, such protease inhibitors strongly bind themselves to specific species of protease and reversibly inhibit their enzymatic activities, fulfilling the function of controlling relevant protease actions. The use of a serine protease inhibitor, therefore, can put a stop to a reaction system that causes a disease. Thus, serine protease inhibitors can be utilized in pharmaceutical preparations.

Nafamostat mesilate is used for alleviating symptoms as abdominalgia and ameliorating hyperamylasemia by virtue of its ability to inhibit the activity of trypsin in the pancreatic duct.

Further, in treating and preventing the thromboses that are symptomatic of disseminated intravascular coagulation syndrome, serine protease inhibitors such as gabexate mesilate and nafamostat mesilate, which inhibit the activities of serine proteases such as activated blood coagulating factor II, activated blood coagulating factor VII, activated blood coagulating factor IX, and activated blood coagulating factor X, have been used.

Gabexate mesilate and nafamostat mesilate, which are conventional serine protease inhibitors, however, also find use also as anti-coagulating agents because these compounds are capable of inhibiting serine protease activities by binding themselves to the active site of serine proteases and of inhibiting the digestive enzymes of chymotrypsins and, at the same time, neutralizing thrombins and Activity Xa. Though the conventional serine protease inhibitors in a sense have been intended to effect inhibition of the activities of serine proteases by binding themselves to the active regions of such serine proteases, they are incapable of attaining high selectivity, depending on the kind of serine protease. The conventional serine protease inhibitors have encountered difficulty in selectively inhibiting only the enzymatic activity of a serine protease taken as a target because the active regions of serine proteases possess comparatively similar structures.

Summary of the Invention

The inventors, after repeating a diligent study on the problems confronting the prior art as described above, have found that a serine protease inhibitor containing a substance which inhibits the reaction of a targeted serine protease with a substrate thereof by binding itself to the substrate of the serine protease in competition with the serine protease is capable of selectively inhibiting only the enzymatic activity of the serine protease. This invention has been perfected based on this knowledge.

This invention comprises the following items (1) – (5).

- (1) A serine protease inhibitor containing a substance which inhibits the reaction of a targeted serine protease with a substrate of the serine protease by binding itself to the substrate of the serine protease in competition with the serine protease.

- (2) An antithrombotic containing the serine protease inhibitor mentioned above.
- (3) An inhibitor against uPA, tPA, and plasmin, containing the serine protease inhibitor mentioned above.
- (4) An agent which resists digestive enzymes, containing the serine protease inhibitor mentioned above.
- (5) A therapeutic agent for disseminated intravascular coagulation, containing the serine protease inhibitor mentioned above.

This invention concerns a highly bioadaptive serine protease inhibitor, which specifically inhibits the activity of a serine protease by binding itself to the substrate of the serine protease in competition with the serine protease. The substance can be an anhydridized serine protease, for example, which binds specifically to the substrate of the serine protease.

Detailed Description of the Invention

The invention relates to a serine protease inhibitor which contains a substance which binds to the substrate of a serine protease in competition therewith, thereby inhibiting the activity of the serine protease. This substance will hereafter be referred to as a "Ser-P inhibiting substance".

The Ser-P inhibiting substance is preferably a substance which possess a structure similar to that of a serine protease such that it is capable of binding the natural substrate thereof and yet does not have the enzymatic activity of that serine protease. The Ser-P inhibiting substance may, for example, be made by substituting, adding, or deleting, by genetic engineering, an amino acid from the relevant serine protease. The modification may, for example, be made by substituting or deleting the active-site serine residue of the relevant serine protease and/or by substituting, adding, or deleting at least one amino acid which is in proximity to the active-site serine residue.

The Ser-P inhibiting substance may also preferably be an anhydridized serine protease. The preferred site of anhydridization is the active-site serine residue where the serine is anhydridized to dehydroalanine. By way of background, it is noted that serine proteases derive their catalytic activity from the hydroxyl group of the active-site serine residue. The inventors have found, however, that the hydroxyl group is not always necessary to confer binding affinity between the serine protease and its natural substrate. Accordingly, if the hydroxyl group is removed in a fashion so as not to create any steric hindrance which would prevent the resulting compound from binding to the natural substrate of the serine protease, the resulting product will still retain binding activity with the substrate but will not have any catalytic activity thereon. Further, since such anhydridized serine proteases are able to bind to the substrate in competition with native serine protease, they serve to inhibit serine protease activity.

In comparison to anhydridized serine proteases, conventional serine protease inhibitors, which are generally low molecular weight compounds, are less specific as they have been engineered to bind the catalytic site of the serine protease.

As all serine proteases have similar catalytic sites, such inhibitors are less specific than anhydridized serine proteases which derive their inhibitory activity by binding specifically to the natural substrate of the serine protease of interest.

Ser-P inhibiting substances may be derived from essentially any serine protease which may be duly anhydridized in a manner such that serine protease activity is eliminated but binding affinity with the natural substrate of the serine protease is retained. Examples of preferred serine proteases from which Ser-P inhibiting substances may be derived include, but are not limited to: activated blood coagulation factors such as activated blood coagulation factor II, activated blood coagulation factor VII, activated blood coagulation factor IX, activated blood coagulation factor X, activated blood coagulation factor XI, and activated blood coagulation factor XII; fibrinolytic enzymes such as uPA, tPA, and plasmin; anti-coagulating proteins such as kallikrein and protein C; and enzymes such as trypsin, chymotrypsin, and acrosin. Especially preferred are Ser-P inhibiting substances derived from activated blood coagulation factor II, activated blood coagulation factor VII, activated blood coagulation factor IX, activated blood coagulation factor X, activated blood coagulation factor XI, and activated blood coagulation factor XII. The source of the serine protease to be anhydridized is not limited by the method used for the production thereof. It may be, for example, obtained by isolation from blood plasma or by genetic engineering.

The serine protease inhibitor of the present invention may include, in addition to the Ser-P inhibiting substance, such known additives such as a stabilizer or an extender in concentrations which do not impair the effect of this invention. It may also contain more than one kind of Ser-P inhibiting substance.

The serine protease inhibitor may be in the form of a powder, a liquid, or particulates.

Anhydridized serine protease is not only useful as a serine protease inhibitor containing a thrombosis preventive of new mechanism but also usable in controlling a serine protease activity in various fields such as the fermentation industry and purification of useful proteins.

Examples of methods for producing an anhydridized serine protease include, but are not limited to:

- (1) A method which comprises sequentially carrying out the following three steps, 1 – 3, in the order mentioned. It is preferred that at least one compound selected from the group consisting of polyhydric alcohols and saccharides be present during the process, at least during the collection step 3., to produce an inhibitor that is capable of retaining a substrate specific binding ability intact.
 1. A step of inducing reaction of the active serine residue site of a serine protease with an inhibitor (1st step)
 2. A step of performing an alkali treatment (2nd step)
 3. A step of performing collection (3rd step)

- (2) A method set forth in method (1) mentioned above, wherein the step for performing the alkali treatment is carried out at a pH value in the range of 11.0 – 13.5.
- (3) A method set forth in method (1) or method (2) mentioned above, wherein at least one compound selected from the group consisting of polyhydric alcohols and saccharides is glycerin, ethylene glycol, or sucrose.
- (4) A method set forth in any of the methods (1) – (3) mentioned above, wherein the salt or amphoteric electrolyte mentioned above is at least one compound selected from the group consisting of sodium chloride, potassium chloride, and glycine.
- (5) A method set forth in any of the methods (1) – (4) mentioned above, wherein at least one compound selected from the group consisting of polyhydric alcohols and saccharides mentioned above accounts for not less than 5% of the whole amount in volume ratio where the compound is in a liquid state or in mass ratio where the compound is in a powdery or solid state at 23°C in an atmosphere having a relative humidity of 50%.
- (6) A method set forth in any of the methods (1) – (5) mentioned above, wherein the concentration of the salt or the amphoteric electrolyte mentioned above is not less than 0.2 M.

Now, the methods for production mentioned above will be described in detail below.

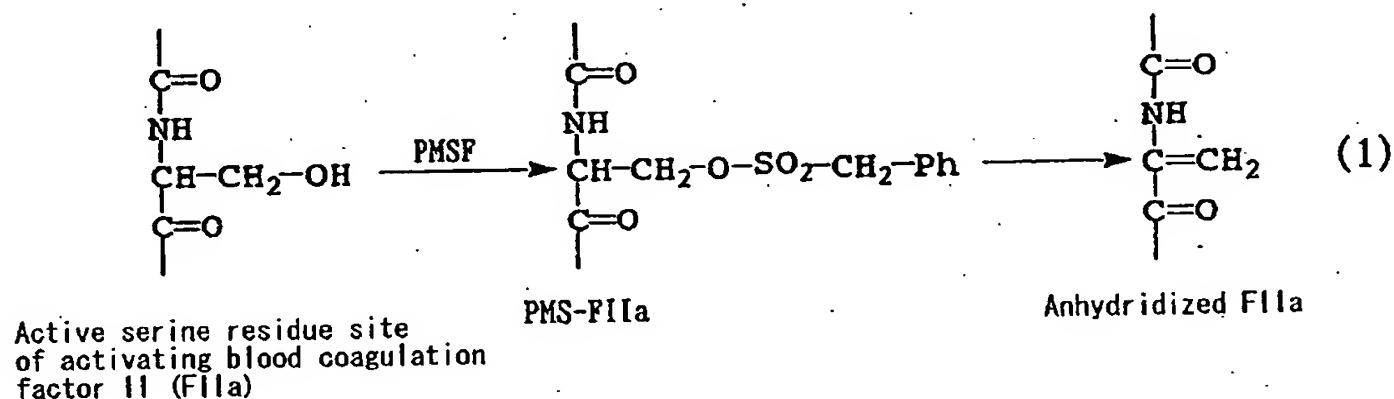
The method for the production of an anhydridized serine protease mentioned above includes

1. A step of inducing reaction of the site of an active serine residue of a serine protease with an inhibitor (1st step)
2. A step of performing an alkali treatment (2nd step)
3. A step of performing collection (3rd step).

The steps mentioned above are executed sequentially in the order mentioned.

This method is characterized by the presence of at least one compound selected from the group consisting of polyhydric alcohols and saccharides, at least during the collection step 3.

The method for the production mentioned above, can be illustrated for example, using an activated blood coagulating factor II as a serine protease and phenylmethanesulfonyl fluoride (PMSF) as an inhibitor, represented by the following reaction formula (I).



FORMULA

The method for producing the anhydridized serine protease mentioned above will be described below by sequentially following the steps 1 – 3 mentioned above.

(1) 1st step

The 1st step consists of causing the site of an active serine residue of a serine protease to react with a synthetic inhibitor and form an ester linkage with the active serine residue of the serine protease and thereby eliminating the serine protease activity.

Examples of the serine protease which may be used include, but are not limited to, activated blood coagulating factor II, activated blood coagulating factor VII, activated blood coagulating factor IX, activated blood coagulating factor X, activated blood coagulating factor XI, and activated blood coagulating factor XII etc., fibrinolytic enzymes, kallicrein, and anti-coagulating proteins.

The synthetic inhibitor used must be capable of forming an ester linkage by the reaction with the active serine residue of a serine protease as illustrated by the reaction formula mentioned above. Examples of suitable synthetic inhibitors include; various kinds of sulfonyl fluoride such as PMSF, 2-phenyl ethane-1-sulfonyl fluoride, methane sulfonyl fluoride, and p-toluene sulfonyl (tosyl) fluoride, tosyl chloride, diisopropyl fluorophosphoric acid (hereinafter referred to occasionally as “DFP”), 3,4-dichloroisocoumarine (hereinafter referred to occasionally as “3,4-DCI”), L-1-chloro-3-[4-tosyl acid]-7-amino-2-heptanone hydrochloric acid (hereinafter referred to occasionally as “TLCK”), and L-1-chloro-3-[4-tosyl acid]-4-phenyl-2-butanone (hereinafter referred to occasionally as “TPCK”).

Such a synthetic inhibitor, prior to the addition thereof to the reaction system, may be dissolved in a solvent such as methanol, acetone, ethanol, propanol, isopropanol, butanol, propan-2-ol, acetone, dimethyl formamide, or dimethyl sulfoxide. Further, the addition of the synthetic inhibitor is preferred to be continued until the activity of the relevant serine protease declines to a level of not more than 3%, preferably not more than 1%, with the decline of the activity to this level attentively pursued meanwhile for the purpose of alleviating the possible trouble of entailing an operation of separating and removing the inhibitor when the addition is excessive and of enhancing the reactivity of the inhibitor.

Further, the reaction solvent may be a salt solution having NaCl incorporated therein for the purpose of adjusting osmotic pressure and ion equilibrium so as to favor the survival of a serine protease or a salt solution formulated in a composition having several species of ion such as K^+ , Ca^{2+} , and Mg^{2+} incorporated therein, to which is added a buffer which can manifest pH values in the range of 2 – 10, preferably in the range of 4 – 8, for the purpose of maintaining a stable pH value as a buffer system. Examples of such buffers include, phosphate buffer, carbonate buffer, bicarbonate buffer, tris-buffer, citric acid-sodium phosphate buffer, succinic acid-sodium hydroxide buffer, potassium phthalate-sodium hydroxide buffer, imidazole-hydrochloric acid buffer, borate buffer physiological saline solution, and Good's buffer, for example.

As regards the reaction conditions, since thermal changes generally have a large effect on the stabilization of a serine protease, the reaction is preferably carried out at a temperature in a range of -30 to $50^{\circ}C$, preferably in the range of 4 to $40^{\circ}C$.

The product obtained by the reaction mentioned above is purified and isolated by using a heretofore known method.

Examples of the method to be used for the purifying and isolating operation include, but are not limited to, gel filtration, ion-exchange chromatography, affinity chromatography, ultrafilter membrane, and dialysis. Typical gel filtration consists in adding the solution obtained after the reaction to a column of gel particles (sephadex, biogel, or agarose, for example) swelled with a solvent and continuously passing the solvent through the column. By this method, first the serine protease product is eluted as a macromolecular solute and subsequently the synthetic inhibitor is eluted as a low molecular solute and, in this manner, the two solutes are separated from each other. The solvent that can be used in the gel filtration may be a salt solution having NaCl incorporated therein for the purpose of adjusting osmotic pressure and ion equilibrium so as to favor the survival of a serine protease or a salt solution formulated in a composition having several species of ion such as K^+ , Ca^{2+} , and Mg^{2+} incorporated therein, which is added a buffer selected from ones which manifest pH values in the range of 2 – 10, preferably in the range of 4 – 8 and for the purpose of maintaining a pH value stably as a buffer system. Examples of the buffer include, phosphate buffer, carbonate buffer, bicarbonate buffer, tris-buffer, citric acid-sodium phosphate buffer, succinic acid-sodium hydroxide buffer, potassium phthalate-sodium hydroxide buffer, imidazole-hydrochloric acid buffer, borate buffer, physiological saline solution, and Good's buffer.

(2) 2nd step and 3rd step

In the 2nd step and 3rd step, for the purpose of dissociating an ester linkage and, at the same time, transforming a serine residue to a dehydroalanine residue thereby producing an anhydridized serine protease and further obtaining the anhydridized serine protease in a high yield by a simple procedure without entailing coagulation and association while the pH is returned from a high pH range to an intermediate pH region for the sake of regeneration, a step of performing an alkali treatment (2nd step)

and a step of performing collection (3rd step) are sequentially carried out in the order mentioned on the product which has been purified and isolated by 1st step. At least one compound selected from the group consisting of polyhydric alcohols and saccharides is present at least during a step for performing collection.

First, the solvent for dissolving the serine protease product purified and isolated at the 1st step can be a salt solution having NaCl incorporated therein for the purpose of adjusting osmotic pressure and ion equilibrium so as to favor the survival of a serine protease or a salt solution formulated in a composition having several species of ion such as K^+ , Ca^{2+} , and Mg^{2+} incorporated therein, which is added a buffer selected from ones which manifest pH values in the range of 2 – 10, preferably in the range of 4 – 8 and for the purpose of maintaining a pH value stably as a buffer system. As concrete examples of the buffer, phosphate buffer, carbonate buffer, bicarbonate buffer, tris-buffer, citric acid-sodium phosphate buffer, succinic acid-sodium hydroxide buffer, potassium phthalate-sodium hydroxide buffer, imidazole-hydrochloric acid buffer, borate buffer, physiological saline solution, and Good's buffer may be cited.

The compound selected from the group consisting of polyhydric alcohols and saccharides and used for the production of an anhydridized serine protease mentioned above is added to be used in combination with a salt or an amphoteric electrolyte for the purpose of promoting the anhydridization without inducing coagulation and association of protein during the alkali treatment in a high pH range and of effecting regeneration of the anhydridized serine protease without inducing coagulation and association during the return of the pH from a high pH range to an intermediate range in the collection. This compound, even when used only in the collection, can fulfill the purpose of producing the anhydridized serine protease mentioned above.

Examples of the compound selected from the group consisting of polyhydric alcohols and saccharides include such polyhydric alcohols which include sugar alcohols as tetritols (such as, for example, erythritol, D-threitol, L-threitol, and D,L-threitol), pentitols (such as, for example, ribitol, D-arabinitol, L-arabinitol, D,L-arabinitol, and xylitol), hexytols (such as, for example, allitol, dulcitol (galactitol), sorbitol (D-glucitol), L-glucitol, D,L-glucitol, D-mannitol, L-mannitol, D,L-mannitol, D-altritol, L-altritol, D,L-altritol, D-iditol, and L-iditol), heptitol, maltitol, lactitol, glycerin, ethylene glycol, diethylene glycol, triethylene glycol, propylene glycol, isopropylene glycol, 1,3-butylene glycol, neopentyl glycol, pentamethylene glycol, hexamethylene glycol, pentaerythritol, dipentaerythritol, tripentaerythritol, trimethylol ethane, trimethylol propane, anhydrous enneahexitol, 1,4-butane diol, 1,2,4-butanetriol, and 1,2,6-hexane triol and such saccharides as glycerin aldehyde dioxyacetone, threose, erythrulose, erythrose, arabinose, ribulose, ribose, xylose, xylulose, lyxose, glucose, fructose, mannose, idose, sorbose, gulose, talose, tagalose, galactose, allose, psicose, altrose, and sucrose may be cited. These compounds may be used either singly or in the form of a mixture of two or more members. Among other compounds enumerated above, at least one compound selected from the group consisting of glycerin, ethylene glycol, and sucrose proves particularly favorable.

The ratio of the at least one compound selected from the group consisting of polyhydric alcohols and saccharides mentioned above to be used as determined under conditions of 23°C in air temperature and 50% in relative humidity is not less than 5%, preferably not less than 15%, based on the whole of the reaction solution in terms of volume ratio in the case of a liquid sample, or in terms of weight ratio in the case of a powdery, particulate, or solid sample. Even when the ratio falls short of 5%, it is made possible to perform the second and the 3rd step infallibly and attain satisfactory manifestation of the effect aimed at by relatively heightening the concentration of the salt or amphoteric electrolyte to be used at the same time. The ratio (i.e. the

concentration) of the at least one compound selected from the group consisting of polyhydric alcohols and saccharides mentioned above to the whole of the reaction solution is preferred to be properly decided at an appropriate level depending on the kind of compound so as to ensure full manifestation of the effect aimed at. For the sake of this decision, it is necessary to consider the kind and the concentration of a salt or an amphoteric electrolyte to be used at the same time.

The salt or the amphoteric electrolyte to be used in the production of an anhydridized serine protease mentioned above is to be used in combination with at least one compound selected from the group consisting of polyhydric alcohols and saccharides for the purpose of promoting the anhydridization without inducing coagulation and association of protein during the alkali treatment in a high pH range and of effecting regeneration of the anhydridized serine protease without inducing coagulation and association during returning of the pH value from a high pH range to an intermediate range in the collection. The salt or the amphoteric electrolyte does not need to be particularly restricted but is only required to be capable of attaining such salt concentration (ionic strength) and such dielectric constant as is suitable for achieving the purpose. It can be either organic or inorganic.

Examples of suitable salt or the amphoteric electrolyte mentioned above include, halogenated alkali metals such as sodium chloride and potassium chloride, hydrogenated alkaline earth metals such as magnesium chloride and calcium chloride, salts of inorganic acids such as ammonium chloride, ammonium sulfate, sodium carbonate, potassium carbonate, magnesium carbonate, ammonium carbonate, calcium carbonate, sodium hydrogen carbonate, calcium hydrogen carbonate, potassium hydrogen carbonate, ammonium hydrogen carbonate, sodium phosphate, disodium hydrogen phosphate, potassium dihydrogen phosphate, diammonium hydrogen phosphate, sodium borate, and potassium borate, salts of organic acids such

as sodium citrate, potassium citrate, magnesium citrate, calcium citrate, ammonium citrate, sodium phthalate, potassium phthalate, magnesium phthalate, calcium phthalate, ammonium phthalate, sodium succinate, potassium succinate, magnesium succinate, calcium succinate, ammonium succinate, sodium acetate, potassium acetate, calcium acetate, magnesium acetate, and ammonium acetate, and amino acids destined to form such amphoteric electrolytes as glycine and alanine. These salts or amphoteric electrolytes may be used either singly or in the form of a mixture of two or more members. Among other salts or amphoteric electrolytes enumerated above, such low molecular alkali metal salts, inorganic salts, and amphoteric electrolytes which are readily soluble in water, easily adjustable to the optimum ionic strength (salt concentration) and dielectrically constant depending on the concentration of the at least one compound selected from the group consisting of polyhydric alcohols and saccharides mentioned above, and easily isolated and purified (such as, for example, dialysis) prove to be particularly advantageous. Specifically, at least one compound selected from the group consisting of sodium chloride, potassium chloride, and glycine is preferred.

The concentration of the salt or amphoteric electrolyte mentioned above is preferably not less than 0.2 M, more preferably not less than 0.5 M. Even when this concentration falls short of 0.2 M, by relatively heightening the proportion of the relevant compound to the whole amount similarly to the case of the at least one compound selected from the group consisting of polyhydric alcohols and saccharides mentioned above, it is made possible to carry out the 2nd and 3rd steps satisfactorily and manifest the expected effect efficiently.

At the 2nd step, to effect the anhydridization by a treatment with an alkali, namely to produce an anhydridized serine protease by subjecting the serine protease product purified and isolated at the 1st step to dissociation of an ester linkage and to

substituting an alanine residue for a serine residue as well, the pH of the reaction system is adjusted by the addition of an alkali to a level of not less than 11.0, preferably to a level in the range of 11.0 – 13.5 and the reaction temperature is retained at a level in the range of 30 to 50 °C, preferably in the range of 4 – 40°C (when necessary, by the addition of at least one compound selected from the group consisting of polyhydric alcohols and saccharides). If the pH mentioned above falls short of 11.0, the shortage will be at a disadvantage in preventing the reaction for removal of PMSF from occurring and the anhydridization from proceeding. As concrete examples of the alkali mentioned above, such monovalent bases as sodium hydroxide and potassium hydroxide, such divalent bases as calcium hydroxide, barium hydroxide, calcium oxide, magnesium oxide, calcium carbonate, and sodium carbonate, and such trivalent bases as iron hydroxide may be cited. If the reaction temperature mentioned above falls short of –30°C, the shortage may result in the disadvantageous freezing the reaction system. If it exceeds 50°C this may result in the undesirable exposure of the serine protease to albuminous degeneration possibly to the extent of rendering infeasible the resumption of the original state even by a subsequent regenerating operation.

Then, at the 3rd step, the solution containing an anhydridized serine protease synthesized by the treatment with an alkali mentioned above is subsequently (after the reaction of anhydridization) is reverted to the original state (the state of stereostructure) by a regenerating operation performed with permitting coexistence of at least one compound selected from the group consisting of polyhydric alcohols and saccharides. The regenerating operation mentioned above may, but does not necessarily have to, utilize a conventionally known method. For example, a method which comprises adjusting the pH of the system (solution) resulting from the reaction with a solvent (the same solvent as used in the reaction of anhydridization mentioned above may be adopted) to a level in the range of 4 – 10 and retaining the system at a

temperature in the range of 30 to 50 °C for a fixed length of time and a method which comprises adjusting the pH by dialysis to a level in the range of 4 – 10, for example.

The serine protease inhibitor of this invention may incorporate therein, in addition to a Ser-P inhibitor, such known additives as stabilizer and extender in concentrations not so high as to impair the effect of this invention.

Further, the serine protease inhibitor of this invention may be in the form of powder, liquid, and particulates.

The serine protease inhibitor of this invention may contain two or more kinds of Ser-P inhibiting substance.

Incidentally, the thrombin is known to bind itself to fibrinogen, factor VIII, factor XIII, and protein C as a substrate and to activate them. The anhydrothrombin which is used in this invention fastens itself to fibrinogen, factor VIII, factor XIII, and protein C respectively at $K_d 4.4 \times 10^{-8}$, $K_d 1.2 \times 10^{-8}$, $K_d 2.8 \times 10^{-7}$, and $K_d 8.1 \times 10^{-5}$. When anhydrothrombin, for example, is added to a blood coagulating system in connection with thrombin, therefore, it is enabled to bind itself to fibrinogen, factor VIII, factor XIII, and protein C and consequently inhibit the activation of these substrates by a thrombin pertinent to the relevant substrate. Even in this case, it is safe to conclude that the substrate specificity is retained intact. This conclusion implies that in view of the actual situation in which the blood coagulating system forms a cascade and promotes the coagulation continuously and, at the same time, the activated substance further activates a factor existing on the upstream side of the reaction system and consequently causes the blood coagulation to advance more quickly, even the inhibitor which originates in a specific serine protease such as, for

example, thrombin is enabled to act synergistically on individual factors and consequently inhibit the blood coagulating system.

Moreover, the initiation of a blood coagulating system is known in two types; i.e. a system initiating a coagulating reaction when blood contacts the surface of an extraneous matter to an organism and a system initiating a coagulating reaction when blood contacts a vital tissue other than the hemangioendothelium. In summary, the coagulating reaction which initially occurs on contact with the surface of an extraneous matter is followed by the activation of the factor XII and the factor XI, further followed by the activation of the factor VII, the factor VIII, and the factor IX, and finally followed by the transformation of prothrombin to thrombin via the activation of the factor X and of fibrinogen to fibrin. In the case of the contact with a vital tissue, the activation of such tissue components as prokallikrein and macromolecular kininogen is consequently followed by the activation of the factor VII, the factor VIII, and the factor IX. The use of an anhydrothrombin, for example, is effective in inhibiting a coagulating system owing to the contact with the surface of an extraneous matter and, at the same time, in inhibiting a coagulating system generated by the contact with a tissue. It inhibitions the activity of a digestive enzyme sparingly. It is further at an advantage of low side effect because it has a structure such that it is decomposed in the system for metabolizing an in vivo substance.

When the Ser-P inhibiting substance is such an anhydro compound of digestive enzyme as trypsinogen, it is effective in alleviating such a subjective symptom as abdominalgia and improving hyperamylasemia by inhibiting activation of trypsinogen.

The serine protease inhibitor of this invention is useful as antithrombotic, antifibrinolytic, agent for resisting a digestive enzyme, and therapeutic agent for disseminated intravascular coagulation.

Particularly, the therapy heretofore used for resisting thrombosis uses such an antivitamin K agent as warfarin but, at the same time, diminishes the production of such a protein as protein C or protein S which functions to resist coagulation. According to the serine protease inhibitor of this invention, however, it is made possible to attain specific inhibition of such a serine protease as causes or participates in abnormal blood coagulation or cancerous growth regarding the therapy and control of the abnormal blood coagulation and the therapy of the cancer. When the anhydro serine protease is actually used as a therapeutic agent, since it resembles a original serine protease thereof in structure excepting the serine residue at the active site, it has the advantage of eluding recognition as a substance extraneous to an organism and of permitting no easy occurrence of an immunological problem.

The mode of administration of the serine protease inhibitor of this invention embraces such injections that are executed intramuscularly, intravenously, etc. When the serine protease which forms the target for inhibition is contained in blood, the inhibitor in question is preferred to be administered by intravenous injection. When the inhibitor is elected to serve the purpose of inhibiting cancerous growth, for example, it may be directly injected into the cancerous tissue.

Further, the inhibitor of this invention as a platelet coagulation inhibitor effectively cures or prevents various diseases caused wholly or partly by the coagulation of blood platelets. Particularly, it is effective as an agent for curing disseminated intravascular coagulation and as a pharmaceutical preparation for inhibiting or preventing such occlusions of blood vessels as myocardial infarction and

cerebral infarction which are caused by thrombosis. It is also effective as an agent for inhibiting acute reocclusion after the surgery for the formation of endermic coronary artery against the thrombosis in the coronary artery during the inhibiting reocclusion with blood platelets emitted during the therapy for thrombus decomposition by using such a thrombolytic agent which performed on the infarcted focus, and further as an agent for inhibiting bloods coagulation during a medical treatment entailing extracorporeal blood circulation.

Further, the platelet coagulation inhibitor of this invention, for use in the therapy of a varying disease, may incorporate therein as occasion demands in addition to the effective components mentioned above, such other components as are effective for pharmaceutical preparations, for example, other kinds of platelet coagulation inhibiting components.

When the inhibitor of this invention is used as an antithrombotic, an antifibrinolytic, an agent for resisting digestive enzyme, or a therapeutic agent against disseminated intravascular coagulation, the clinical dose thereof given by injection is preferably in the range of 1 – 100000 mg, more preferably in the range of 10 – 50000 mg, and particularly preferably in the range of 100 – 1000 mg as reduced to the effective component mentioned above per adult, though depending on the amount of an enzyme taken as the target for inhibition and the degree of inhibition. The administration of this dose does not need to be limited to once daily. Optionally, the dose may be split into a plurality of fractions and given individually on as many separate occasions within a day. The dose may be given by instillation, depending on the kind of disease and the seriousness of disease. The platelet coagulation inhibitor of this invention mentioned above may be administered once daily. Otherwise, the dose may be split into two or three portions, which are individually given on separated occasions. When the compound of this invention is used for extracorporeal

circulation, this use may be effected in the mode using an injection mentioned above. The dose involved in this case follows the example of the dose of the injection mentioned above.

Best Mode of Embodying the Invention

Now, this invention will be specifically described below with reference to examples.

Example 1

To a solution formed by dissolving 5.21 mg of an activated blood coagulating factor II (α -thrombin) derived from Cohn Paste III in 10 ml of a 5mM phosphate buffer/0.1 M NaCl/pH 6.5, 30 μ l of a 7% phenylmethanesulfonyl fluoride (PMSF) methanol solution was added at intervals of 30 minutes until the total activity reached a level of less than 1%. The produced solution was cooled to 0°C and the cooled solution and 0.5 ml of 1 M NaOH subsequently added thereto were left to react with each other for 12 minutes. To the resultant reaction solution, 5 ml of a 3M NaCl solution was added and then 19 g of glycerin was further added.

The solution consequently formed was adjusted to pH 7.8 by the use of a 1 M tris hydrochloride buffer/50% glycerin/pH 7 and then retained at 4°C for 12 hours. Thereafter, it was dialyzed against a 50mM tris hydrochloride buffer/1 M NaCl/pH 7.5 and further against a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 and then treated with p-amidinophenylmethanesulfonyl fluoride (made by Wako Pure Chemical Industries, Ltd; hereinafter referred to simply as "APMSF") until the residual activity was completely inactivated. The resultant solution was added to a benzamidine sepharose column equilibrated with a 50 mM tris hydrochloride

buffer/0.1 M NaCl/pH 7.5. The impurity peak was completely washed with the same buffer and the adsorbate was eluted with a 50 mM tris hydrochloride buffer/0.1 M NaCl/0.2 M benzamideine/pH 7.5. The eluted solution was dialyzed against a 50 mM tris hydrochloride buffer/ 1 M NaCl/pH 7.5 to remove the benzamidine and obtain 2.8 mg of an anhydridized activating blood coagulating factor II (anhydrothrombin).

Example 2

To a solution formed by dissolving 2.5 mg of an activated blood coagulating factor VII derived from human blood plasma in a 10 ml of a 5 mM phosphate buffer/0.1 M NaCl/pH 6.4, 30 μ l of a 7% phenylmethanesulfonyl fluoride (PMSF) methanol solution was added at intervals of 30 minutes until the total activity reached a level of less than 1%. The produces solution was cooled to 0°C and the cooled solution and 0.5 ml of 1 M NaOH subsequently added thereto were left to react with each other for 12 minutes. To the resultant reaction solution, 5 ml of a 3 M NaCl solution was added and then 19 g of glycerin was further added.

The solution consequently formed was adjusted to ph 7.8 by the use of a 1 M tris hydrochloride buffer/50% glycerin/pH 7 and then retained a 4°C for 12 hours. Thereafter, it was dialyzed against a 50 mM tris hydrochloride buffer/1 M NaCl/pH 7.5 and further against a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 and then treated with APMSF until the residual activity was completely inactivated. The resultant solution was added to a benzamidine sepharose column equilibrated with a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5. The impurity peak was completely washed with the same buffer and the adsorbate was eluted with a 50 mM tris hydrochloride buffer/0.1 M NaCl/0.2 M benzamideine/pH 7.5. The eluted solution was dialyzed against a 50 mM tris hydrochloride buffer/1 M NaCl/pH 7.5 to

remove the benzamidine and obtain 1.2 mg of an anhydridized activating blood coagulating factor VII.

Example 3

To a solution formed by dissolving 4.0 mg of an activated blood coagulating factor IX derived from human blood plasma in 10 ml of a 5 mM phosphate buffer/0.1 M NaCl/pH 6.5, 30 μ l of a 7% phynylmethanesulfonyl fluoride (PMSF) methanol solution was added at intervals of 30 minutes until the total activity reached a level of less than 0.1%.

The produced solution was cooled to 0°C and the cooled solution and 0.5 ml of 1 M NaOH subsequently added thereto were left to react with each other for 12 minutes. To the resultant reaction solution, 5 ml of a 3 M NaCl solution was added and then 19 g of glycerin was further added.

The solution consequently formed was adjusted to pH 8 by the use of a 1 M tris hydrochloride buffer/pH 7 and then left standing and thereafter dialyzed against a 50 mM tris hydrochloride buffer/1 M NaCl/pH 7.5 at 4°C for 12 hours. It was further dialyzed thereafter against a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 at 4°C for 12 hours and then treated with APMSF until the residual activity was completely inactivated. The resultant solution was added to a benzamidine sepharose 6B column equilibrated with a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 at 4°C and washed unadsorbed component completely with the same buffer. Then the adsorbate was eluted with a 50 mM tris hydrochloride buffer/0.1 M benzamidine/0.1 M NaCl/pH 7.5.

The eluted solution was dialyzed to remove the benzamidine and obtain about 2.1 mg of an anhydridized activating blood coagulating factor IX.

Example 4

To a solution formed by dissolving 3.2 mg of an activated blood coagulating factor X derived from human blood plasma in 10 ml of a 5 mM phosphate buffer/0.1 M NaCl/pH 6.5, 30 μ l of a 7% phynylmethanesulfonyl fluoride (PMSF) methanol solution was added at intervals of 30 minutes until the total activity reached a level of less than 0.1%.

The produced solution was cooled to 0°C and the cooled solution and 0.5 ml of 1 M NaOH subsequently added thereto were left to react with each other for 12 minutes. To the resultant reaction solution, 5 ml of a 3 M NaCl solution was added and then 19 g of glycerin was further added.

The solution consequently formed was adjusted to pH 8 by the use of a 1 M tris hydrochloride buffer/pH 7 and then left standing at 4°C for 12 hours and thereafter dialyzed against a 50 mM tris hydrochloride buffer/1 M NaCl/pH 7.5 at 4°C for 12 hours. It was further dialyzed thereafter against a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 at 4°C for 12 hours and then treated with APMSF until the residual activity was completely inactivated. The resultant solution was added to a benzamidine sepharose 6B column equilibrated with a 50 mM tris hydrochloride buffer/0.1 M NaCl/pH 7.5 at 4°C and washed unadsorbed component completely with the same buffer. Then the adsorbate was eluted with a 50 mM tris hydrochloride buffer/0.1 M benzamidine/0.1 M NaCl/pH 7.5.

The eluted solution was dialyzed to remove the benzamidine and obtain about 1.4 mg of an anhydridized activating coagulating factor X.

Example 5

A sample formed by adding the anhydridized activated blood coagulating factor II produced in Example 1 to 20 ml of human whole blood at a ratio of 5 mg/ 1 ml (5mM, Tris-HCl, 0.15 M NaCl, pH 7.5) was tested for coagulation time with a thromboelastogram. The coagulation time consequently found was about three times as much as that obtained of a sample formed without addition of the anhydridized activating blood coagulating factor II (1 ml, 5 mM Tris-HCl, 0.15 M NaCl, ph 7.5 added).

Example 6

A sample formed by adding the anhydridized activated blood coagulating factor VII produced in Example 2 to 20 ml of human whole blood at a ratio of 0.5 mg/ 1 ml (5mM, Tris-HCl, 0.15 M NaCl, pH 7.5) was tested for coagulation time with a thromboelastogram. The coagulation time consequently found was about 2.1 times as much as that obtained of a sample formed without addition of the anhydridized activating blood coagulating factor VII (1 ml, 5 mM Tris-HCl, 0.15 M NaClL, ph 7.5 added).

Example 7

A sample formed by adding the anhydridized activated blood coagulating factor IX produced in Example 3 to 20 ml of human whole blood at a ratio of 0.5 mg/ 1 ml (5mM, Tris-HCl, 0.15 M NaCl, pH 7.5) was tested for coagulation time with a

thromboelastogram. The coagulation time consequently found was about 3 times as much as that obtained of a sample formed without addition of the anhydridized activating blood coagulating factor IX (1 ml, 5 mM Tris-HCl, 0.15 M NaCl, ph 7.5 added).

Example 8

A sample formed by adding the anhydridized activated blood coagulating factor X produced in Example 4 to 20 ml of human whole blood at a ratio of 0.5 mg/1 ml (5mM, Tris-HCl, 0.15 M NaCl, pH 7.5) was tested for coagulation time with a thromboelastogram. The coagulation time consequently found was about 1.8 times as much as that obtained of a sample formed without the addition of the anhydridized activating blood coagulating factor X (1 ml, 5 mM Tris-HCl, 0.15 M NaCl, ph 7.5 added).

Industrial Applicability

The serine protease inhibitor of this invention is capable of selectively inhibiting exclusively the enzymatic activity of a serine protease aimed at. Thus, it is not only useful as a thrombosis preventive of a new mechanism but also usable efficiently for the control of a serine protease activity in varying fields such as fermentation industry and purification of usable proteins.